TORTUOSIDE, A NEW NATURAL COUMARIN GLUCOSIDE FROM SESELI TORTUOSUM

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ABSTRACT.—A new coumarin glucoside, tortuoside [3], has been isolated from the aerial parts of *Seeli tortuosum*. Its structure was established on the basis of spectroscopic data and chemical evidence.

During the course of our investigations on metabolites from Seseli tortuosum L.B.S. Eur. (Umbelliferae), we have isolated and characterized, in addition to previously reported compounds (1,2), three coumarin derivatives. Two of these, (-)-2'-senecioyloxy-1',2'-dihydroxanthyletin [1] and (-)-2'-isovaleryloxy-1'-2'-dihydroxanthyletin [2], have been reported from various species of Seseli (3,4) while the third, named tortuoside [3], is described here.

Chromatographic procedures applied to the Me₂CO extract of the air-dried aerial parts of S. tortuosum provided tortuoside [3], $C_{20}H_{26}O_{10}$, mp 212–216°, $[\alpha]D + 18.9^{\circ}$. An analysis of its ¹H-nmr spectrum clearly indicated that 3 contains one 7,8-disubstituted coumarin nucleus and a β -glycoside unit, as evidenced from the large ¹H coupling constant (7.6 Hz) of the anomeric proton (δ 4.56). The carbon resonances of 3 were in agreement with a β -D-glucosyl residue (5). However, the chemical shift of the anomeric carbon (δ 98.2) seemed anomalous. This upfield shift from the expected values was indicative of a tert-O- β -D-glucopyranoside (6,7).

The aglycone moiety, isolated from the hydrolysis of **3**, was identified as 7hydroxy-8-(2',3'-dihydroxy-3'-methylbutyl)-coumarin [**4**], on analytical and spectroscopical bases (1). The ¹³C, ¹H-COSY of **4** led to the full clarification of the carbon and proton signals (Table 1 and Experimental) and corroborated the assigned structure. The sugar isolated as the methylglycoside was identical in all respects with an authentic sample of methyl- α -D-glucoside. Placement of this residue at the tertiary carbon on the aglycone was confirmed by comparison of the chemical shifts of the C-3', C-4', and C-5' in **3** with those of the corresponding signals of **4** (6). ¹³C-nmr data of acetates **5** and **6** were consistent structure for these compounds. Further elution



- 4 R=H, R'=H, R''=H
- 5 R=Ac, R'=Ac, R"= β -D-glucosyl acetate
- **6** R = Ac, R' = Ac, R'' = H
- 7 R=Me, R'=H, R''=H

Carbon	Compound						
	1	2	3	4	5 ⁶	6 ^c	7
C-2	160.9 113.0 142.9 112.6 128.5 115.9 156.3 140.4 154.1 27.7 69.0	160.9 113.1 142.9 112.6 128.1 115.8 156.2 104.3 154.1 27.7 69.1	163.9 111.9 146.5 113.5 128.3 114.3 155.1 115.5 161.1 26.3 79.3	163.8 111.9 146.5 113.4 128.2 114.2 155.1 115.5 161.3 26.4 79.4 74 9	159.9 115.5 143.2 116.3 126.3 118.8 152.1 ^d 119.3 153.5 ^d 23.6 79.3 77.2	160.2 115.4 143.6 116.4 126.4 118.9 152.0 ^d 119.5 153.3 ^d 23.9 78.2 71.8	163.8 112.7 143.9 113.0 126.8 107.4 160.5 115.8 153.3 25.9 78.0 72.8
C-3'	76.6 24.9 27.2 165.5 115.4 158.0 20.1 22.9	76.4 24.9 27.2 171.8 43.2 25.5 22.0 22.1	81.4 22.9 ^d 22.9 ^d 98.2 75.3 78.2 ^e 71.7 78.1 ^e 62.8	74.0 25.3 ^d 25.5 ^d	77.2 23.6 ^e 24.3 ^e 95.4 72.9 71.7 ^f 68.9 71.4 ^f 62.4	71.8 25.1 ^e 25.3 ^e	72.8 24.0 ^d 25.4 ^d 56.1

TABLE 1. ¹³C nmr Data.^a

^aCompounds 1, 2, 5–7 in CDCl₃; compounds 3 and 4 in CD₃OD.

^bMeCO: 168.6, 170.3, 170.1, 169.8, 169.3, 168.9; CH₃CO: 20.3, 20.4, 20.5, 20.7, 21.7.

^{d-f}Signals in any vertical column with the same superscript may be reversed.

with CHCl₃-MeOH (9:1) gave 0.9 g of tortuoside [**3**]: $[\alpha]D + 18.9^{\circ}$ (c = 0.01, MeOH), mp 212–216°; ¹H nmr (CD₃OD) δ 1.38 (3H, s, 3'-Me), 1.42 (3H, s, 3'-Me), 2.58–3.89 (10H, m, H-1', H-2', and glucose-H), 4.56 (1H, d, anomeric-H), 6.18 (1H, d, J = 9.4 Hz, H-3), 6.82 (1H, d, J = 8.5 Hz, H-6), 7.88 (1H, d, J = 9.4 Hz, H-4); ¹³C nmr see Table 1. *Anal.* calcd for C₂₀H₂₆O₁₀, C 56.33, H 6.15; found C 56.12, H 6.25.

HYDROLYSIS OF TORTUOSIDE [3].— A solution of 0.25 g of tortuoside [3] in 10 ml of 10% methanolic HCl was refluxed under magnetic stirring for 4 h. After being cooled, the reaction mixture was neutralized with a NaHCO₃ solution, diluted with H₂O, and extracted with CHCl₃. The combined organic layers were washed with H₂O, dried, and evaporated. Chromatography of the residue and elution with CHCl₃-MeOH (95:5) gave 0.15 g of aglycone 4: $[\alpha]D + 60.6^{\circ}$ (c = 0.009, MeOH); mp 138-140°; ¹³C nmr see Table 1. Anal. calcd for C₁₄H₁₆O₅, C 63.63, H 6.10; found C 63.48, H 6.17.

The aqueous phase was lyophilized to give a methyl- α -D-glycoside identical in all respects with an authentic sample of methyl- α -D-glucoside.

ACETYLATION OF TORTUOSIDE [3].— A solution of 0.1 g of the glucoside 1 and 2 ml of Ac₂O in 6 ml of C₅H₅N was kept at room temperature for 12 h. After usual workup, the crude product was chromatographed. Elution with CHCl₃ gave 0.12 g of acetate 5: ¹H nmr (CDCl₃) δ 1.30 (3H, s, 3'-Me), 1.40 (3H, s, 3'-Me), 1.81 (3H, s, 2'-OAc), 2.01, 2.03, 2.04, 2.07 (12H, each s, OAc-glucose), 2.45 (3H, s, 7-OAc), 3.04 (1H, dd, J = 7.8, 14.6 Hz, H-1'), 3.16 (1H, dd, J = 2.4, 14.6 Hz, H-1'), 3.60–5.35 (8H, m, H-2' and glucose-

^cMeCO: 168.6, 170.5, CH₃CO: 20.7, 20.5.

H), 6.38 (1H, d, J = 9.5 Hz, H-3), 7.04 (1H, d, J = 8.4 Hz, H-5), with the proposed structures. The absolute stereochemistry of the new natural glycoside was determined by comparing the optical rotation value of the methyl derivative 7 with that of natural meranzin hydrate (8). Consequently, tortuoside [3] is expressed as 7-hydroxy-8-[(2'*R*)hydroxy-3'-O- β -D-glucopyranosyl-3'methylbutyl]-coumarin.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.— Melting points were taken on a Reichert micro hotstage and are uncorrected. Elemental analyses were carried out on a Carlo Erba Model 1106 Elemental Analyzer. All nmr experiments were performed on a Bruker AC 200 spectrometer with 5mm ¹H and ¹³C probes operating at 200 and 50 MHz respectively. All ¹H-nmr and ¹³C-nmr chemical shifts were referred to internal TMS. Cc was carried out on 0.063–0.200 mesh Merck Si gel. All extracts were dried over Na₂SO₄.

PLANT MATERIAL.—Plant material was collected in September 1987, near Perugia, Umbria, Italy, and voucher specimens (Number 2315) were deposited in the Herbarium of the Dipartimento di Biologia Vegetale of the University of Perugia.

EXTRACTION AND ISOLATION OF THE COM-PONENTS.—Dried and finely powdered *S. tortuosum* aerial parts (2 kg) were extracted exhaustively with Me₂CO. The resulting extracts were concentrated under vacuum. The crude gum was chromatographed over Si gel, and elution with CHCl₃ gave, in addition to previously reported compounds (1), 1.5 g of **1** and 0.3 g of **2** (2,3), whose ¹³C-nmr data are presented for the first time (Table 1), and fully supported the 7.38 (1H, d, J = 8.4 Hz, H-6), 7.69 (1H, d, J = 9.5 Hz, H-4); ¹³C nmr see Table 1. *Anal.* calcd for C₃₂H₄₄O₁₆, C 56.14, H 6.48; found C 54.24, H 6.36.

ACETYLATION OF 4.—A solution of 50 mg of aglycone 4 and 1 ml of Ac₂O in 3 ml of C₅H₅N was kept at room temperature for 12 h. After usual workup, the crude product was chromatographed. Elution with CHCl₃ afforded 54 mg of acetate 6: ¹H nmr (CDCl₃) δ 1.29 (3H, s, 3'- Me), 1.31 (3H, s, 3'-Me), 1.83 (3H, s, 2'-OAc), 2.38 (3H, s, 7-OAc), 3.09 (1H, dd, J = 10.3, 14 Hz, H-1'), 3.20 (1H, dd, J = 2.4, 14 Hz, H-1'), 5.17 (1H, dd, J = 2.4, 10.3 Hz, H-2'), 6.40 (1H, d, J = 9.4 Hz, H-3), 7.04 (1H, d, J = 8.5Hz, H-5), 7.38 (1H, d, J = 8.5 Hz, H-6), 7.72 (1H, d, J = 9.4 Hz, H-4); ¹³C nmr see Table 1. *Anal.* calcd for C₁₈H₂₀O₇, C 62.06, H 5.79; found C 62.22, H 5.62.

METHYLATION OF 4.—To a solution of 4 (75 mg) in Me₂CO (5 ml) were added 67 mg of K₂CO₃ and 0.13 ml of MeI under N₂ and magnetic stirring. The solution was refluxed for 2 h. After cooling, the mixture was diluted with H₂O and extracted with CHCl₃. The combined organic layers were washed with H₂O, dried, and evaporated. Chromatography of the residue and elution with CHCl₃-MeOH (95:5) gave 61 mg of (+)-meranzin hydrate [7]: $[\alpha]D + 51.2^{\circ}$ (c = 0.01, EtOH) [lit. (8) (-)-meranzin hydrate: $[\alpha]D - 57^{\circ}$].

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